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1Hydroxypropyl methylcellulose-beeswax edible coatings formulated with
2antifungal food additives to reduce alternaria black spot and maintain
3postharvest quality of cold-stored cherry tomatoes

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27**Abstract**

28Edible composite coatings based on hydroxypropyl methylcellulose (HPMC), beeswax
29(BW), and food preservatives with antifungal properties were formulated and evaluated
30on cherry tomatoes during cold storage. Selected food preservatives included: sodium
31methyl paraben (SMP), sodium ethyl paraben (SEP) and sodium benzoate (SB).
32Cherry tomatoes artificially inoculated with *Alternaria alternata* were coated and stored
33up to 21 d at 5 °C followed by 4 d of shelf-life at 20 °C. All antifungal coatings reduced
34the incidence and severity of alternaria black spot on inoculated cherry tomatoes, being
35the SB-based coating the most effective. Analytical and sensory fruit quality was
36evaluated on intact and cold-stored tomatoes. In contrast to coatings containing SMP
37or SEP, the SB-based coating was effective to reduce weight loss and respiration rate
38and maintain the firmness of coated cherry tomatoes. Peel color, ethanol and
39acetaldehyde content of the juice, sensory flavor, off-flavors, and fruit appearance were
40not adversely affected by the application of the antifungal coatings. In conclusion,
41HPMC-BW coatings containing the food additive SB at 2% showed potential for
42industrial application, including the production and commercialization of organic cherry
43tomatoes.

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45**Keywords:** *Solanum lycopersicum*; edible coatings; hydroxypropyl methylcellulose;
46postharvest quality; food preservatives; *Alternaria alternata*

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511. Introduction

52 Tomato fruits have a relatively short postharvest life and during fruit ripening
53 many processes reducing fruit quality may take place, leading to important economic
54 losses. Therefore, the development of new technologies to effectively control ripening
55 and decay would be of great economic importance (Hoeberichts et al., 2002).
56 *Alternaria alternata* (Fr.) Keissl., causing black spot, is among the most common fungal
57 pathogens responsible for postharvest decay of cherry tomato fruit (Wang et al., 2008).
58 The use of synthetic chemical fungicides as antimicrobial agents to control fungal
59 spoilage of fresh horticultural products has been practiced for many years. However,
60 concerns about environmental contamination and human health risks associated with
61 fungicide residues on/in produce, as well as the proliferation of fungicide-resistant
62 strains of the pathogens, have led to serious restrictions or even bans of many
63 synthetic fungicides (Palou et al., 2008). At present, there is a lack of authorized
64 postharvest treatments and/or registered fungicides available for the control of
65 postharvest diseases of high value commercial fruits, such as tomato. Alternative
66 methods that have been proposed for the control of postharvest diseases include
67 biological control with antagonistic microorganisms, physical methods such as heat or
68 radiations, and the use of low-toxicity chemicals with antimicrobial activity (Montesinos-
69 Herrero et al., 2009; Palou et al., 2002, 2008; Valencia-Chamorro et al., 2009a). The
70 latest include natural or synthetic compounds of known and low toxicity, usually
71 classified as safe food-grade additives or Generally Regarded as Safe (GRAS)
72 substances by international authorities (Larrigaudière et al., 2002; Palou et al., 2002).

73 In recent years, the release of antimicrobial agents incorporated into
74 biodegradable edible films and coatings has emerged as a new, effective, and
75 environmentally-friendly alternative mean to extend the shelf-life of many products
76 including fresh fruits and vegetables. Edible coatings provide a semi-permeable barrier
77 to water vapor, oxygen (O₂), and carbon dioxide (CO₂) that reduce weight loss and
78 respiration. Additional advantages of edible coatings are the possibility to maintain the

79firmness of the fruit and provide gloss to coated products (Greener-Donhowe and
80Fennema, 1994). Edible coatings are based on polysaccharides, proteins and lipids or
81a mixture of these. Other food-grade ingredients such as antimicrobial agents,
82antioxidants, flavors, color pigments, and vitamins can also be incorporated into the
83basic formulation of these coatings with the aim to improve their functional properties
84(Valencia-Chamorro et al., 2011a). Among the active ingredients used in antimicrobial
85edible coatings, compounds such as plant essential oils, food aromas, organic acids,
86parabens, their salts and other permitted food additives or GRAS compounds, have
87been preferred for fruit and vegetables (Das et al., 2013; Fagundes et al., 2013;
88Valencia-Chamorro et al., 2009a; Xu et al., 2007). Our research group optimized stand-
89alone hydroxypropyl methylcellulose (HPMC)-lipid edible composite films containing a
90wide variety of food additives and GRAS compounds such as mineral salts, organic
91acid salts and their mixtures, and sodium salts of parabens and their mixtures to
92provide antifungal activity against the citrus pathogens *Penicillium digitatum* and *P.*
93*italicum* (Valencia-Chamorro et al., 2008). Then, selected coatings were tested *in vivo*
94against green and blue molds on different citrus cultivars. The inhibitory activity of the
95coatings was strongly dependent on the susceptibility of each citrus cultivar to
96penicillium decay and the storage temperature (Valencia-Chamorro et al., 2009a,
972009b, 2010, 2011b). Similar studies also proved the antifungal activity of several
98mineral salts, organic acid salts, and paraben salts incorporated to HPMC-BW coatings
99against the pathogens *Monilinia fructicola* in artificially inoculated plums (Karaca et al.,
1002014) and *Botrytis cinerea* and *A. alternata* in inoculated cherry tomatoes during shelf-
101life at 20 °C (Fagundes et al., 2013). In a recent work, the best coatings against *B.*
102*cinerea* were evaluated on cherry tomatoes cold-stored at 5 °C and it was observed
103that the effect of the coatings on disease development and fruit quality during storage
104was dependent on the storage temperature, remarking the need to evaluate the
105coatings under commercial storage conditions (Fagundes et al., 2014). In our previous
106study to select appropriate antifungal coatings for the control of alternaria black spot of

107cherry tomato, the best results after incubation of coated fruit at 20 °C were obtained
108with HPMC-BW coatings containing 2.0% sodium benzoate (SB), sodium ethyl paraben
109(SEP), or sodium methyl paraben (SMP) (Fagundes et al., 2013). The objective of the
110present research was to determine the effect of selected HPMC-BW edible coatings
111formulated with antifungal food additives on the development of alternaria black spot
112and the physico-chemical and sensory quality of cherry tomatoes during cold storage.
113This information is needed for potential commercial development of suitable antifungal
114edible coatings.

115

1162. Materials and methods

1172.1. Materials

118 HPMC (Methocel E15) was purchased from Dow Chemical Co. (Midland, MI,
119USA). BW (grade 1) was supplied by Fomesa Fruitech, S.L. (Beniparrell, València,
120Spain). Oleic acid and glycerol were from Panreac Química, S.A (Barcelona, Spain).
121Laboratory reagent grade preservatives (99% minimum purity) were purchased from
122Fluka Chemie AG (Buchs, Switzerland) and Merck KGaA (Darmstadt, Germany), and
123included SMP ($C_8H_7NaO_3$; E-218), SEP ($C_9H_9NaO_3$; E-214), and SB ($C_7H_5O_2Na$; E-211).
124All these chemicals are classified as food additives (with their correspondent E-
125number) or GRAS compounds by the European Food Safety Authority (EFSA) and the
126United States Food and Drug Administration (US FDA).

127

1282.2. Emulsions preparation

129 HPMC-lipid edible composite emulsions were prepared combining the
130hydrophilic phase (HPMC) and the hydrophobic phase (BW) suspended in water.
131Glycerol and oleic acid were used as plasticizer and emulsifier, respectively. All the
132formulations contained 30% BW (dry basis, db) and the ratios of HPMC-glycerol (3:1)
133(db) and BW-oleic acid (5:1) (db) were kept constant throughout the study. Tween 80
134was also added to the formulations at a concentration of 1.5% (w/w) to improve wetting

of the coating and adherence to the tomato fruit. All formulations contained 2.0% (w/w) of food preservative. Emulsions were prepared as described by Valencia-Chamorro et al. (2008). Briefly, an aqueous solution of HPMC (5% w/w) was prepared by dispersing the HPMC in hot water at 90 °C and later hydration at 20 °C. The corresponding food preservative, BW, glycerol, oleic acid, and water were added to the HPMC solution and heated at 98 °C to melt the lipids. Samples were homogenized with a high-shear probe mixer (Ultra-Turrax model T25, IKA-Werke, Steufen, Germany) for 1 min at 12.000 and 3 min at 22.000 rpm. Emulsions were cooled under agitation to a temperature lower than 25 °C by placing them in a water bath and agitation was continued during 25 min to ensure complete hydration of the HPMC. The emulsions were prepared with a final solid concentration of 10% and had a viscosity in the range of 140-147 cp. Table 1 shows the viscosity and pH of the emulsions containing selected food preservatives. Emulsions were kept 1 d at 5 °C before use. These formulations were stable and no phase separation was observed.

149

2.3. Effect of coatings on disease development

2.3.1. Fungal inoculum

The strain TAV-6 of *A. alternata*, obtained from decayed tomato fruit in Valencia packinghouses, was isolated, identified, and maintained in the IVIA culture collection of postharvest pathogens. Prior to each experiment, the isolate was grown on potato dextrose agar (PDA; Sigma-Aldrich Chemie, Steinheim, Germany) in petri dishes at 25 °C for 7-14 d. From this culture, a high-density conidial suspension was prepared in Tween 80 (0.05%, w/v; Panreac-Química S.A., Barcelona, Spain) and sterile water. This suspension was passed through two layers of cheesecloth, measured with a haemocytometer, and diluted with sterile water to achieve an inoculum density of 1×10^6 spores/mL of *A. alternata*.

161

2.3.2. Fruit inoculation and coating application

Cherry tomatoes (*Solanum lycopersicum* L. var. *cerasiforme* cv. Josefina; syn.: *Lycopersicon esculentum* Mill.) used in the experiments were commercially grown and collected in the Valencia area (Spain) and stored up to 24 h at 5 °C until use. Fruit were free from previous postharvest treatments or coatings. Before each experiment, fruit were selected, randomized, washed with fruit biodegradable detergent at 6% (v/v) (Essasol V., Didsa, Potries, Valencia), rinsed with tap water, and allowed to air-dry at room temperature. Cherry tomatoes were superficially wounded once in the equator with a stainless steel rod with a probe tip 1 mm wide and 2 mm in length. This wound was inoculated with the pathogen by placing 10 µl of a spore suspension containing 1×10^6 spores/ml of *A. alternata*. After incubation at 20 °C for 24 h, inoculated fruit were coated by immersion for 30 s in the selected HPMC-BW edible composite emulsions, drained, and allowed to air-dry at 20 °C. Inoculated but uncoated fruit were used as controls. Coated fruit were placed on plastic trays on corrugated cartons and stored up to 21 d at 5 °C and 90-95% RH, followed by 4 d of shelf-life at 20 °C. In every experiment, each treatment was applied to 3 replicates of 10 fruit each. The experiments were repeated twice.

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2.3.3. Determination of disease incidence and severity

The incidence of alternaria black spot was calculated as the percentage of decayed fruit. Disease severity was determined as the diameter of the lesion (mm). Incidence and severity were assessed after 7, 14 and 21 d during the storage period at 5 °C, and also after the shelf-life period of 4 d at 20 °C.

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2.4. Effect of coatings on fruit quality

2.4.1. Fruit coating and storage

For the quality study, before each experiment, fruit were selected, randomized, washed with fruit biodegradable detergent, rinsed with tap water, and allowed to air-dry at room temperature. Intact and healthy fruit were divided into four groups of 120 fruit

each, which corresponded to the three coating treatments and one control (uncoated fruit). Cherry tomatoes were coated as described above, drained of excess coating, dried and stored for up to 15 d at 5 °C and 90-95 % RH. Physico-chemical and sensory fruit quality was assessed initially and after 10 and 15 d at 5 °C plus a shelf-life period of 5 d at 20 °C.

2.4.2. *Assessment of fruit quality*

2.4.2.1 *Internal quality*

The assessed internal quality attributes were soluble solids content (SSC), titratable acidity (TA), and pH of tomato juice. SSC of the juice was measured using a digital refractometer (model PR1; Atago Co. Ltd., Japan) and values were expressed as percentage. TA of tomato juice was determined by titrating 5 mL of juice sample with 0.1 M sodium hydroxide to an end point of pH 8.1 and expressed as g of citric acid per 1 L. pH of the juice was determined using a pH-meter (model C830, Consort bvba, Turnhout, Belgium). For each treatment, 3 juice samples from 7 fruit each were prepared and three different readings were performed.

2.4.2.2. *Weight loss*

Lots of 30 fruit per treatment were used to measure weight loss. The same marked cherry tomato were weighed at the beginning and at the end of each storage period. The results were expressed as the percentage of initial weight lost.

2.4.2.3. *Peel color*

Skin color of cherry tomatoes was measured with a Minolta (Model CR-400, Minolta, Tokyo, Japan) on 20 fruit per treatment, using the CIE (Commission Internationale de l'Eclairage) color parameters lightness (L^*), a^* , b^* , chroma (C^*) and hue angle (h°). Each measurement was taken at three locations for each cherry tomato. A standard white calibration plate was employed to calibrate the colorimeter.

219

2202.4.2.4. *Fruit firmness*

221 Firmness of 20 fruit per treatment was determined at the end of each storage
222period using an Instron Universal testing machine (Model 4301, Instron Corp., Canton,
223MA, USA). Each fruit was compressed between two flat surfaces closing together at
224the rate of 5 mm/min. The machine gave the deformation (mm) after application of a
225load of 9.8 N to the equatorial region of the fruit. Results were expressed as
226percentage of deformation, related to initial diameter.

227

2282.4.2.5. *Respiration rate*

229 Respiration of coated and uncoated cherry tomatoes was measured by the
230closed system. Three replicates of 5 fruit each were used to determine the CO₂
231production at the end of the storage period. Samples were weighed and placed in
232sealed containers of known volume. The accumulation of CO₂ in the headspace
233atmosphere was measured at 20 °C over a period of 3 h. The gas sample (1 mL) was
234injected into a gas chromatograph (GC) (Thermo Trace, Thermo Fisher Scientific, Inc.
235Waltham, MA, USA) equipped with a thermal conductivity detector (TCD) and fitted
236with a Poropack QS80/100 column (1.2 m × 0.32 cm i.d.). Temperatures were 35, 115,
237and 150 °C, respectively for the oven, injector, and thermal conductivity detector.
238Helium was used as carrier gas at a flow rate of 22 mL/min. The CO₂ concentration
239was calculated using the peak area obtained from a standard gas mixture of 15.0:2.5%
240O₂:CO₂. Results were expressed as mg CO₂/kg h.

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2422.4.2.6. *Ethanol and acetaldehyde contents*

243 Ethanol and acetaldehyde were analyzed from the head-space of tomato juice
244from samples using a GC (Thermo Trace, Thermo Fisher Scientific) equipped with an
245auto-sampler (Model HS 2000), flame ionization detector (FID), and 1.2 m x 0.32 cm
246(i.d.) Poropack QS 80/100 column. The injector was set at 175 °C, the column at 150

247°C, the detector at 200 °C, and the carrier gas at 28 mL/min. A composite juice of three
248replicates of ten fruit per treatment was analyzed. Five mL of juice were transferred to
24910-mL vials with crimp top caps and TFE/silicone septum seals. Samples were frozen
250and stored at –18 °C until analyses. A 1-mL sample of the headspace was withdrawn
251from vials previously equilibrated in a water bath at 20 °C for 1 h, followed by 15 min at
25240 °C, to reach equilibrium in the headspace, and then injected into the GC. Ethanol
253and acetaldehyde was identified by comparison of retention times with standards.
254Results were expressed as mg of gas per 1 L of juice.

255

2562.4.2.7. *Sensory evaluation*

257 Sensory quality of treated samples was evaluated by 10 trained judges at the
258end of each storage period and shelf-life (ISO8586-1:1993). Each judge was given
259samples from each batch and requested to evaluate flavor on a 9-point scale, where 1
260= very poor and 9 = optimum, and off-flavor on a 5-point scale, where 0 = absence of
261off-flavor and 5 = high presence of off-flavor. Ten fruit per treatment were halved cut
262and separated into individual segments. Two segments from two different fruit were
263presented to judges in trays labeled with 3-digit random codes and served to them at
264room temperature. The judges had to taste the segments of each sample in order to
265compensate, as far as possible, for biological variation of the material. Spring water
266was provided for palate rinsing between samples. External aspect of the fruit (coating
267cracks, spots, etc.) was also evaluated by the panelists. A 3 point scale was used in
268which the aspect was classified as 1 = bad, 2 = acceptable, and 3 = good. Panelists
269were also asked to rank visually the coated fruit from highest to lowest gloss.

270

2712.5. *Statistical analysis*

272 Statistical analysis was performed using Statgraphics 5.1. (Manugistics Inc.,
273Rockville, MD, USA). Specific differences between means were determined by Fisher's
274protected least significant difference test (LSD, $P < 0.05$) applied after an analysis of

275variance (ANOVA). For sensory gloss, specific differences were determined by
276Friedman test, which is recommended for ranking by the UNE 87023 (AENOR, 1997).
277For disease incidence data, the ANOVA was applied to the arcsine of the square root
278of the percentage of infected fruit in order to assure the homogeneity of variances.
279Non-transformed means are shown.

280

2813. Results and discussion

2823.1. Effect of coatings on disease development

283 During cold storage at 5 °C, the assayed coatings significantly reduced
284alternaria black spot incidence and severity compared with uncoated samples, except
285the SMP coating that after 14 d at 5 °C was not effective to reduce disease incidence
286(Fig. 1). After 7 d at 5 °C, control samples showed 100% disease incidence, whereas
287all the coatings completely inhibited mold growth. In general, the reduction of disease
288severity by coating application was considerably higher than the reduction of disease
289incidence. At the end of the 21-d cold storage period, the SEP and SB coatings were
290the most effective to reduce the severity of black spot (reduction of around 65%).
291These coatings also reduced disease incidence effectively, being those containing SB
292the most effective (reduction of around 70% and 30% after 14 and 21 d of storage,
293respectively). When tomatoes were transferred to 20 °C to simulate shelf-life, the
294coatings did not prevent the onset of disease, and black spot incidence basically
295reached 100%. This result might have been influenced by the high concentration of
296fungal inoculum that was used in these trials (10^6 spores/mL). This high inoculum
297density of *A. alternata* was used to obtain high percentages of decay on control fruit
298and to conservatively select only those coatings with the highest potential for effective
299commercial usage. From results of disease incidence and severity, it can be confirmed
300that the activity of edible coatings was fungistatic rather than fungicidal, because mold
301growth was delayed but not completely inhibited and both incidence and severity
302increased with storage time. In general, comparable differences in performance

303depending on the amount of fungal inoculum, fruit cultivars or fruit characteristics have
304been observed with most of the alternative antifungal treatments, which mode of action
305is rather fungistatic than fungicidal (Palou et al., 2008).

306 Among the three food additives tested as antifungal coating ingredients, SB was
307the most effective against *A. alternata*, followed by SEP, whereas SMP lost
308effectiveness after 14 d at 5 °C (Fig. 1). SB is among the most widely used
309antimicrobial food additives. Its antimicrobial activity is pH-dependent, being the
310undissociated form the most effective. Therefore, the use of this additive is
311recommended in acidic products such as citrus fruits, and good disease control
312performance has been reported with SB applied both as aqueous solution (Palou et al.,
3132002) and as a fruit coating ingredient (Valencia-Chamorro et al., 2010). Parabens and
314their sodium salts are GRAS compounds with increasing interest as alternative low-
315toxicity means to control postharvest decay in fresh horticultural products. For instance,
316satisfactory control of citrus postharvest diseases was observed when SMP and SEP
317were included in the formulation of antifungal edible coatings (Valencia-Chamorro et
318al., 2009a) and also when they were applied as postharvest dip treatments (Moscoso-
319Ramírez et al., 2013a, 2013b). Parabens are in the undissociated form at pH values of
320most foods ($pK_a = 8.5$) and are effective over a wide pH range of 4–8 (Thompson,
3211994). Paraben salts like SMP and SEP are more soluble in water than their
322correspondent parabens and they might interfere on both the germinative and
323vegetative phases of microbial development, although it has been reported that fungal
324spore germination is much more susceptible than vegetative growth (Watanabe and
325Takesue, 1976). It has been suggested that the general mode of action of these salts is
326through an uncoupling of oxidative phosphorylation, inhibition of NAD^+ and FAD-linked
327mitochondrial respiration, or the reduction of mitochondrial membrane potential (Soni et
328al., 2001).

329

3303.2. *Effect of coatings on fruit quality*

3313.2.1. *Fruit internal quality*

332 The antimicrobial HPMC-BW coatings did not significantly affect SSC, TA and
333pH of cherry tomatoes (Table 2). During storage and shelf-life, SSC increased and TA
334decreased with respect to the values at harvest, as a consequence of an increase in
335the content of soluble sugars and a decrease in the amount of organic acids. In
336general, the effect of coating application on fruit internal quality parameters has been
337proven to be dependent on coating type, fruit cultivar and storage conditions. While
338some authors found no differences in these parameters after coating application on
339different citrus cultivars (Baldwin et al., 1995; Obenland et al., 2008), others observed
340lower SSC and TA reductions when compared to uncoated controls, which was always
341related to decreases in weight loss and respiration rate (Togrul and Arslan, 2004). In
342some works with tomatoes, the application of edible coatings resulted in lower SSC (Ali
343et al., 2010; Yaman and Bayoindirli, 2002) and TA (Das et al., 2013) than in uncoated
344samples, which was attributed to the fact that the coatings provided a semi-permeable
345barrier to gases around the fruit, modifying the internal atmosphere by reducing O₂ and/
346or elevating CO₂ and suppressing ethylene production.

347

3483.2.2. *Weight loss*

349 Fig. 2 shows the weight loss on coated and uncoated samples stored for 10 and
35015 d at 5 °C, followed by 5 d of shelf-life at 20 °C. Tomatoes are naturally covered by a
351continuous wax layer that provides high resistance to water movement across the
352cuticle. Coatings containing hydrophobic compounds, deposited as an additional layer
353over the natural waxes, should improve the moisture resistance of the fruit (Fagundes
354et al., 2014). In our work, the barrier properties of the coatings were greatly influenced
355by the different food additives incorporated to the HPMC-BW matrix. Whereas SB
356slightly but significantly reduced tomato weight loss ($P \leq 0.05$), SEP and SMP
357increased it compared to uncoated samples. This might indicate a partial removal of
358the natural waxes present on the peel or a negative interaction of these particular salts

359with the waxes. According to the literature, edible coating application has significant or
360not significant effects on weight loss of fruits depending on intrinsic characteristics of
361both the coating and the fruit. Thus, for example, the addition of lipids to
362polysaccharides did not reduce weight loss of coated commodities, such as cherries or
363cucumbers (Baldwin et al., 1997), apples (Bai et al., 2002), or plums (Navarro-
364Tarazaga et al., 2008). In addition, several works confirmed that the addition of food
365preservatives to HPMC-based coatings greatly affects the moisture barrier properties of
366stand-alone films or coatings when applied to different fruits such as citrus, table
367grapes or cherry tomatoes (Fagundes et al., 2014; Pastor et al., 2011; Valencia-
368Chamorro et al., 2008, 2009a). Furthermore, weight loss after the application of some
369HPMC-based coatings clearly depended on the commodity and cultivar. For instance,
370HPMC-lipid coatings containing organic acids salts and their mixtures significantly
371reduced weight loss of coated 'Clemenules' and 'Ortanique' mandarins during long-
372term storage, but did not reduced that of 'Valencia' oranges, and some coatings even
373induced a significant increase in weight loss of this cultivar (Valencia-Chamorro et al.,
3742009b, 2010, 2011b). In previous research with cherry tomatoes, the addition of
375sodium propionate and potassium carbonate to similar HPMC-based coatings also
376increased weight loss compared to uncoated samples (Fagundes et al., 2014). This
377was correlated with the intrinsic moisture barrier of the coating, but also with the
378mechanical properties that might affect coating integrity during prolonged storage of the
379fruit. Compared to the properties of stand-alone films, coating performance is affected
380by coating distribution over the fruit surface, especially whether it forms a continuous
381layer or penetrates into pores. Moreover, fruit skin morphology (presence of hairs,
382thickness and type of cuticle, number of stomata, lenticels, and even cracks in the
383lenticels) and coating physical properties such as surface tension and viscosity strongly
384influence mass transfer of the coated fruit (Hagenmaier and Baker, 1993). For these
385reasons, beyond *in vitro* determination of film properties, the evaluation of coating

386performance in *in vivo* trials with target commodities is mandatory to assess the actual
387potential for industrial application of novel fruit coatings.

388

3893.2.3. *Peel color*

390 The color change that accompanies maturation in many fruits is one of the most
391important quality criteria used by growers and consumers to judge the harvest time and
392the commercial quality. Thus, in tomatoes, the red color is the most visible and
393important quality attribute for marketing. This color is the result of a combination of
394carotenoid pigments, being lycopene (red) the most abundant, followed by carotenes
395(yellow to orange) and xanthophylls (yellow) (López et al., 2001). Table 2 shows the
396CIE color parameters of coated and uncoated cherry tomatoes after 15 d of storage at
3975 °C plus 5 d at 20 °C. In uncoated samples, there was a decrease in L^* , b^* and h°
398values with storage time. At the end of storage, the coatings helped to maintain b^* and
399 h° values, whereas coated tomatoes presented lower L^* than uncoated ones and no
400differences were observed in a^* values among treatments. The differences in b^* values
401between coated and uncoated tomatoes translated into lower h° (which indicates more
402reddish tonalities) and a slight decrease in C^* (purity or saturation of a single color) in
403uncoated than in coated samples. Several works reported a delay in color changes in
404tomatoes during storage at 20 °C by coating application related to its capacity to create
405a modified atmosphere in the fruit (Ali et al., 2010; Zapata et al., 2008; Zhuang and
406Huang, 2003). However, we reported in previous research (Fagundes et al., 2014) that
407although the use of HPMC-BW coatings with different food preservatives reduced
408respiration rate in cherry tomatoes, their effect was insufficient to produce a significant
409change in the peel color of coated cherry tomatoes during cold storage. Furthermore,
410the initial full-developed red color of the cherry tomatoes used in this study (Table 2)
411could also explain the small changes observed in color during cold storage and the
412shelf-life storage period of 5 d at 20 °C.

413

4143.2.4. *Fruit firmness*

415 Firmness values, expressed as percentage of deformation, of coated and
416uncoated samples are shown in Fig 3. No significant differences were observed
417between cherry tomatoes coated with the SB-based coating and the uncoated control,
418whereas the samples treated with the SMP and SEP-based coatings presented higher
419deformation values and consequently lower firmness than the control. The effect of
420coatings on the maintenance of fruit firmness is usually related to their control of weight
421loss and/or the modification of the internal atmosphere of the fruit (Baldwin et al., 1997;
422Seymour et al., 1993; Yaman and Bayoindirli, 2002). Thus, firmness retention in coated
423tomato has been repeatedly related to a reduction in enzymatic activities caused by a
424modification of the internal atmosphere of the fruit (Ahmed et al., 2013; Ali et al., 2010;
425Park et al., 1994; Zapata et al., 2008; Zhuang and Huang, 2003). In this work, cherry
426tomatoes coated with SEP-based HPMC-BW coatings had higher respiration rate than
427control and the rest of coated samples during storage (Fig. 4), which could be related
428with lower fruit firmness. However, this relationship was not observed for SMP-coated
429samples. In any case, the samples with the lowest firmness (fruit treated with coatings
430containing SMP and SEP) suffered the highest weight loss (Figs. 2, 3).

431

4323.2.5. *Respiration rate*

433 The effect of the coatings on respiration rate of cherry tomatoes during cold
434storage plus 5 d at 20 °C is shown in Fig. 4. In general, there was an increase in the
435respiration rate of cherry tomatoes during storage, which indicates an increase in the
436fruit metabolic activity. Ideally, the effect of coatings on respiration of horticultural
437products is related to their ability to create a barrier to O₂ diffusion through the coating,
438which translates in lower respiration rates in coated fruits (Fagundes et al., 2014).
439However, the capacity to create this O₂ barrier greatly depends on the amount and
440nature of minor ingredients in the coating formulation. Additional ingredients, such as
441antioxidant or antimicrobial food additives, can modify their effectiveness (Valencia-

Chamorro et al., 2008). In this work, only the tomatoes coated with the SB-based coatings showed lower respiration rates than uncoated samples, whereas SEP-coated tomatoes had the highest respiration rates. Valencia-Chamorro et al. (2008) reported a 10-fold increase in O₂ permeability of HPMC-lipid edible films amended with SEP compared to SB-based films, which could explain the differences in respiration rate induced by these formulations when applied to cherry tomatoes.

3.2.6. *Ethanol and acetaldehyde content*

The application of HPMC-BW coatings increased ethanol content in the juice of coated cherry tomatoes ($P < 0.05$; Fig. 5). Overall, the concentrations of acetaldehyde and ethanol in the juice of coated cherry tomatoes after both storage periods were in the range of 0.76-1.30 mg/L and 4.2-13.09 mg/L, respectively, while they were in the range of 0.43-0.55 mg/L and 0.32-0.66 mg/L, respectively, in uncoated samples. These values were higher than those reported in previous work by Fagundes et al. (2014), when similar HPMC-based coatings amended with different antimicrobial agents were applied to cherry tomatoes, showing the importance of the role that minor ingredients may play in the final performance of edible coatings. In general, the effect of the application of coatings amended with SMP or SEP on quality parameters like weight loss, respiration rate, and firmness indicates that these antifungal additives affected negatively the metabolism of cherry tomatoes, accelerating the metabolic activity and increasing the production of volatile compounds. However, as it will be discussed in the next subsection, the increment of these volatiles in the juice of cherry tomatoes was not high enough to induce noticeable off-flavors in coated fruit.

3.2.7. *Sensory evaluation*

The HPMC-BW based coatings containing food preservatives did not modify the flavor of cherry tomatoes compared to uncoated samples. The panelists considered the flavor as acceptable irrespective of the treatments and the storage time. At the end of

the storage period, after 15 d of storage at 5 °C plus 5 d at 20 °C of shelf-life, flavor scores were around 5.6-6.4 (considered as acceptable) and no differences were detected among coated and uncoated samples (data not shown). In this study, the panelists detected a slight off-flavor after storage (0.6-0.9) but no differences between coated and uncoated samples were observed, which indicates that it was not due to coating application but to storage time. It can be deducted from these results that the ethanol and acetaldehyde levels reached after cold storage at 5 °C plus 5 d at 20 °C were below the threshold of off-flavors detection for cherry tomatoes. Ali et al. (2010) reported that flavor and overall acceptability of tomatoes coated with gum arabic depended on the solid content of the formulation. Thus, only tomatoes coated with 10% gum showed the highest scores in all parameters after 20 d of storage, while tomatoes coated with 15 or 20% gum presented off-flavors and were not acceptable by the panel of experts.

In this work, the addition of selected food preservatives to HPMC-BW emulsion resulted in stable emulsions. After storage, all coated samples were evaluated as acceptable, although fruit coated with the SEP-based coating were rated as those with the best external appearance after 15 d at 5 °C plus 5 d of shelf-life at 20 °C (data not shown). After both storage periods, none of the tested coatings provided higher gloss than the uncoated control. Similar results were reported by Fagundes et al. (2014) working with HPMC-BW coatings amended with other antifungal compounds like potassium carbonate and sodium propionate that had been selected for their antifungal activity against *B. cinerea*, the cause of tomato postharvest gray mold. This behavior was related to the macroemulsion character of coating formulations (Hagenmaier and Baker, 1994).

4. Conclusion

During cold storage at 5 °C, all the coatings reduced black spot incidence and severity compared with uncoated samples, and the SB-based coating was the most

498effective at inhibiting the development of *A. alternata*. When tomatoes were transferred
499to 20 °C to simulate shelf-life, the coatings did not prevent the onset of disease in these
500artificially inoculated fruit, but significantly reduced disease severity. The SB-based
501coating was also effective to reduce weight loss and respiration rate of cherry
502tomatoes, whereas the sodium salts of paraben tested negatively affected these quality
503parameters. None of the antifungal coatings affected negatively the color, sensory
504flavor, off-flavor, and fruit appearance. Overall, HPMC-BW edible composite coatings
505containing SB as antifungal agent could be a promising industrial treatment to control
506black spot and maintain the postharvest quality of tomatoes destined to both domestic
507or export markets. Since SB is a common food additive, the use of these edible
508coatings could be also of interest for organic producers. Further research should focus
509on the improvement of the physical characteristics of these HPMC-BW edible
510composite coatings in order to increase water loss control and enhance gloss and
511visual quality of coated fruit. Moreover, future work should also consider testing the
512selected SB coating on other cultivars and types of tomato fruits in order to widen its
513potential commercial usage.

514

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656**Table 1.** Characteristics of hydroxypropyl methylcellulose-beeswax edible composite coatings

657containing antifungal food preservatives

Food preservative	E-number	Molecular formula	Solid concentration (%)	Viscosity (cp)	pH
Sodium methyl paraben	E-218	C ₈ H ₇ NaO ₃	10.0	140.4	9.60
Sodium ethyl paraben	E-214	C ₉ H ₉ NaO ₃	10.0	147.0	9.70
Sodium benzoate	E-211	C ₇ H ₅ O ₂ Na	10.0	142.3	6.39

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664**Table 2.** Soluble solid content (SSC), titratable acidity (TA), pH and peel color (CIE parameters)

665of cherry tomatoes coated with hydroxypropyl methylcellulose-beeswax edible composite

666coatings containing antifungal food preservatives and stored at 5 °C for 15 d followed by 5 d of

667shelf-life at 20 °C.

Food preservative	Quality attributes							
	SSC		TA					
	pH	(%)	(g citric acid/L)	<i>L</i> [*]	<i>a</i> [*]	<i>b</i> [*]	<i>C</i> [*]	<i>h</i> [°]
Control ^a	4.47 a	7.27 a	4.05 ab	35.60 a	15.23 a	18.04 a	23.63 a	49.86 a
Sodium methyl paraben	4.47 a	7.15 a	4.04 ab	33.96 b	14.89 a	19.59 b	24.64 ab	52.83 b
Sodium ethyl paraben	4.45 a	6.77 a	4.15 b	34.48 b	14.63 a	19.86 b	24.73 b	53.75 b
Sodium benzoate	4.54 b	7.07 a	3.84 a	34.38 b	15.00 a	19.47 b	24.63 ab	52.54 b

668Values at harvest: TA = 4.63 g citric acid/L; SSC = 6.48%; pH = 4.32; *L*^{*} = 36.85; *a*^{*} = 14.47;669*b*^{*} = 19.32; *C*^{*} = 22.71; *h*[°] = 54.10670^a Control = uncoated.

671Means in columns with different letters are significantly different according to Fisher's protected

672LSD test (*P* < 0.05) applied after an ANOVA.

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677**Figure captions**

678**Fig. 1.** Disease incidence and severity (\pm SD) of black spot on cherry tomatoes
679artificially inoculated with *Alternaria alternata*, uncoated (Control) or coated 24 h later
680with hydroxypropyl methylcellulose-beeswax edible composite coatings containing:
681sodium methyl paraben (SMP), sodium ethyl paraben (SEP) or sodium benzoate (SB),
682stored at 5 °C for up to 21 d followed by 4 d of shelf-life at 20 °C. For each storage
683period, columns with different letters are significantly different by Fisher's protected
684LSD test ($P < 0.05$) applied after an ANOVA. For disease incidence, the ANOVA was
685applied to arcsine-transformed values. Non-transformed means are shown.

686

687**Fig. 2.** Weight loss of cherry tomatoes uncoated (Control) or coated with hydroxypropyl
688methylcellulose-beeswax edible composite coatings containing sodium methyl paraben
689(SMP), sodium ethyl paraben (SEP) or sodium benzoate (SB), stored at 5 °C for 10 or
69015 d followed by 5 d of shelf-life at 20 °C. For each storage period, columns with
691different letters are different by Fisher's protected LSD test ($P < 0.05$) applied after an
692ANOVA.

693

694**Fig. 3.** Firmness of cherry tomatoes uncoated (Control) or coated with hydroxypropyl
695methylcellulose-beeswax edible composite coatings containing: sodium methyl
696paraben (SMP), sodium ethyl paraben (SEP) or sodium benzoate (SB), stored at 5 °C
697for 10 or 15 d followed by 5 d of shelf-life at 20 °C. For each storage period, columns
698with different letters are different by Fisher's protected LSD test ($P < 0.05$) applied after
699an ANOVA.

700

701**Fig. 4.** Respiration rate of cherry tomatoes uncoated (Control) or coated with
702hydroxypropyl methylcellulose-beeswax edible composite coatings containing, sodium
703methyl paraben (SMP), sodium ethyl paraben (SEP) or sodium benzoate (SB), stored
704at 5 °C for 10 or 15 d followed by 5 d of shelf-life at 20 °C. For each storage period,

705columns with different letters are different by Fisher's protected LSD test ($P < 0.05$)
706applied after an ANOVA.

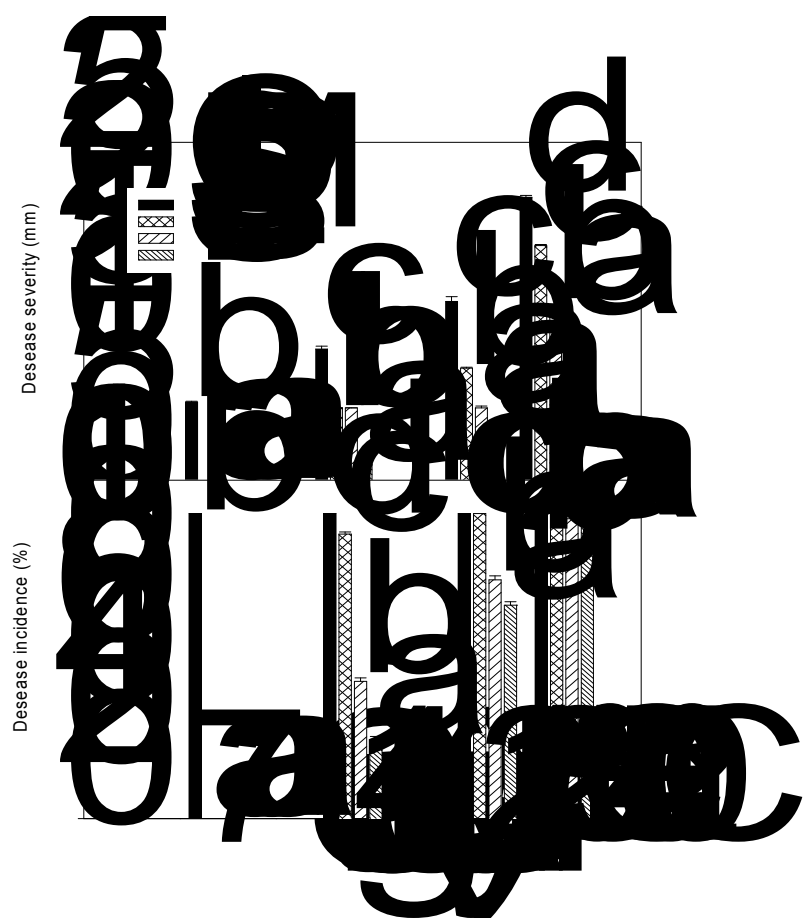
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708**Fig. 5.** Ethanol and acetaldehyde content in the juice of cherry tomatoes uncoated
709(Control) or coated with hydroxypropyl methylcellulose-beeswax edible composite
710coatings containing sodium methyl paraben (SMP), sodium ethyl paraben (SEP) or
711sodium benzoate (SB), stored at 5 °C for 10 or 15 d followed by 5 d of shelf-life at 20
712°C. For each storage period, columns with different letters are different by Fisher's
713protected LSD test ($P < 0.05$) applied after an ANOVA.

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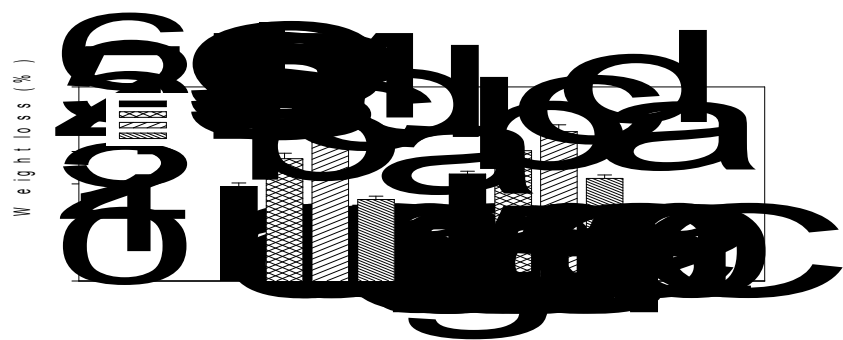
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Figure 1.

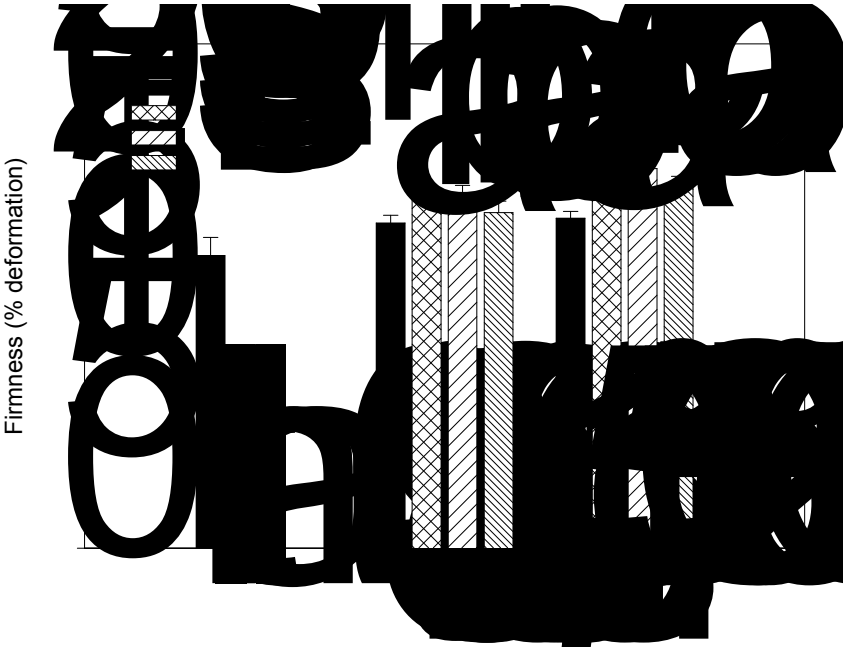
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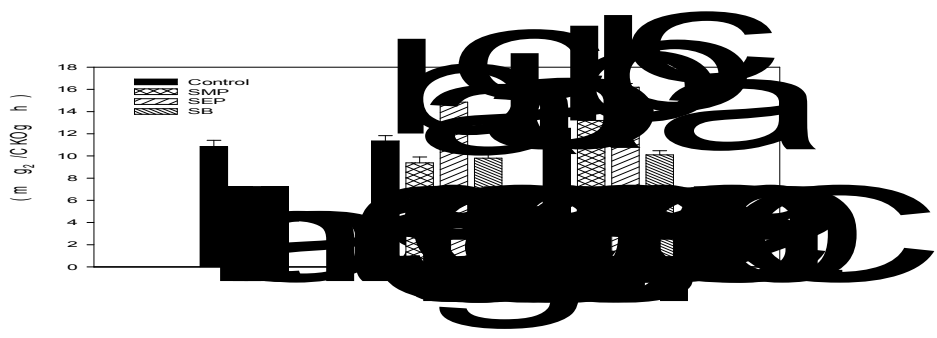
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Figure 3.

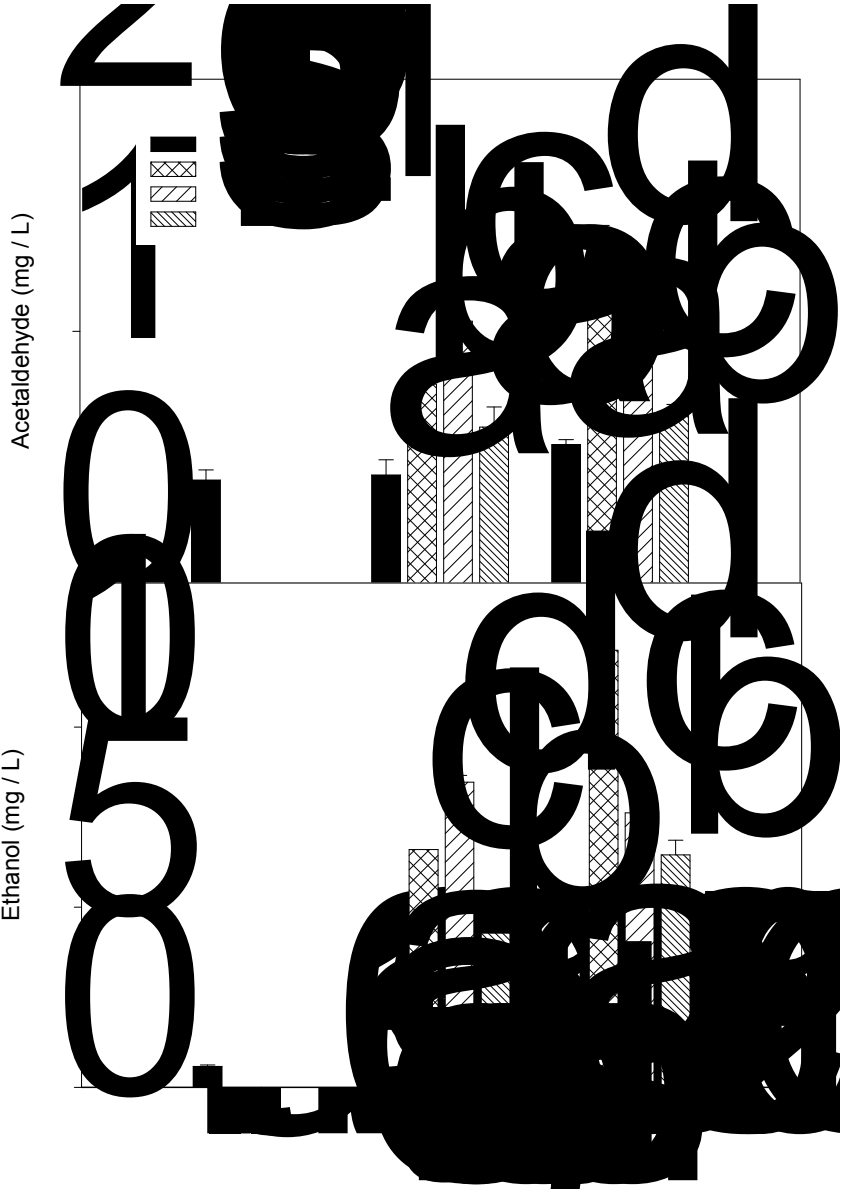
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Figure 4.

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Figure 5.